





# **Final Report**

Report Number: SDWH-M201905389-2(E)

# Skin Sensitization Test of Nerve and Muscle Stimulator

According to ISO 10993-10:2010
Guinea Pig Maximization Test
0.9% Sodium Chloride Injection Extract

Sponsor: Shenzhen XFT Medical Limited

Room203, Building 1, Biomedicine Innovations Industrial Address: Park, #14 Jinhui Road, Pingshan New District, Shenzhen,

China



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# **Supplementary Explanation**

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- (1) Please apply for rechecking within 15 days of receiving the report if there are any objections.
- (2) Any erasure or without special inspection and testing seal renders the report null and void.
- (3) The report is only valid when signed by the persons who edited, checked and approved it.
- (4) The results relate only to the articles tested.
- (5) The report shall not be reproduced except in full without the written approval of the institute.

# **Verification Dates**

Report No.: SDWH-M201905389-2(E)

Test Article Receipt	2019-12-12
<b>Protocol Effective Date</b>	2019-12-19
<b>Technical Initiation Date</b>	2019-12-20
<b>Technical Completion Date</b>	2020-01-16
Final Report Completion Date	2020-02-27

Edited by: Chennyrong 2020-02-25

Date

Reviewed by: Zhang Yan 2020-02-27

Study Director Date

Approved by: Wang 1 Jie 2020-02-27

Authorized Signatory Date

Sanitation & Environment Technology Institute, Soochow University

## Summary

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## 1 Test Article

<b>Test Article Name</b>	Nerve and Muscle Stimulator
Manufacturer	Shenzhen XFT Medical Limited
Address	Room203, Building 1, Biomedicine Innovations Industrial Park, #14 Jinhui Road, Pingshan New District, Shenzhen, China
Model	XFT-2001E
Lot/Batch	Not supplied by sponsor (N/S)

## 2 Main Reference

ISO 10993-10:2010 Biological evaluation of medical devices — Part 10: Tests for irritation and skin sensitization

## 3 Test Method

Potential skin sensitization of test article was evaluated using guinea pig maximization test in accordance with ISO 10993-10:2010 Biological evaluation of medical devices — Part 10: Tests for irritation and skin sensitization.

Study protocol number: SDWH-PROTOCOL -M201905389-2.

## 4 Conclusion

Under the conditions of this study, the test article Nerve and Muscle Stimulator extract showed no significant evidence of causing skin sensitization in the guinea pig. The positive rate of sensitization was 0%. No evidence of skin sensitization in guinea pigs was found.

## **Test Report**

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## 1 Purpose

The test was designed to evaluate the potential of a test article to cause skin sensitization. The test is used as a procedure for screening of contact allergens in guinea pigs and extrapolating the results to humans, but it does not establish the actual risk of sensitization.

## 2 Reference

ISO 10993-10:2010 Biological evaluation of medical devices — Part 10: Tests for irritation and skin sensitization

ISO 10993-12:2012 Biological evaluation of medical devices — Part 12: Sample preparation and reference materials

ISO 10993-2:2006 Biological evaluation of medical devices — Part 2: Animal welfare requirements

## 3 Compliance

ISO/IEC 17025:2005 General requirements for the competence of testing and calibration laboratories (CNAS—CL01 Accreditation criteria for the competence of testing and calibration laboratories) China National Accreditation Service for Conformity Assessment LABORATORY ACCREDITATION CERTIFICATE Registration No. CNAS L2954.

RB/T 214—2017 Competence assessment for inspection body and laboratory mandatory approval—General requirements for inspection body and laboratory Certification and Accreditation Administration of the People's Republic of China INSPECTION BODY AND LABORATORY MANDATORY APPROVAL Certificate No. CMA 180015144061.

## 4 Identification of Test and Control Articles

#### 4.1 Test Article

Test Article Name	Nerve and Muscle Stimulator						
Manufacturer	Shenzhen XFT Medical Limited						
Address	Room203, Building 1, Biomedicine Innovations Industrial Park, #14						
	Jinhui Road, Pingshan New District, Shenzhen, China						
Test Article Initial	Not Sterilized						
State							
CAS Code	N/S						
Model	XFT-2001E						
Size	N/S						
Lot/Batch	N/S						
Test Article Material	N/S						
Packaging Material	N/S						
Physical State	Solid						
Color	N/S						
Density	N/S						
Stability	N/S						
Solubility	N/S						
Storage Condition	Room Temperature						
Intended Clinical Use	This device is intended to address the lack of ankle dorsiflexion in						
	patients who have sustained damage to upper motor neurons or						
	pathways to the spinal cord.						

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The information about the test article was supplied by the sponsor wherever applicable.

#### 4.2 Control Article

## 4.2.1 Negative Control

Article Name: 0.9% Sodium Chloride Injection (SC) Manufacturer: Guangxi Yuyuan Pharmaceutical Co., Ltd.

Size: 500mL

Lot/ Batch#: H19082605 Physical State: Liquid Color: Colorless

Storage Condition: Room Temperature

#### **4.2.2 Positive Control**

Article Name: 2, 4-Dinitrochlorobenzene (DNCB)

Manufacturer: Chengdu Aikeda Chemical Reagent Co., Ltd.

Size: 100g

Lot/ Batch#: 201904101 Induction Concentration: 0.5% Challenge Concentration: 0.1%

Solvent: 0.9% Sodium Chloride Injection

Date prepared: 2019-12-02 Physical State: Liquid Color: Light Yellow

Storage Condition: Room Temperature

## 5 Equipment and Reagents

## 5.1 Equipment

Equipment Name	Equipment Number	Calibration Expire
Horizontal Large Capacity Constant Temperature Vibrator	SDWH897	2020-05-04
Vertical pressure steam sterilizer	SDWH2097	2020-04-16
Steel straight scale	SDWH463	2020-07-29
Electronic scale	SDWH442	2020-05-04

## 5.2 Reagents

Reagent Name	Manufacturer	LOT
Freund's adjuvant, complete liquid	SIGMA	SLBX3240
Sodium dodecyl sulfate (SDS)	Sinopharm Chemical Reagent Co., Ltd	20181210

## 6 Identification of Test System

Species: Hartley guinea pig (Cavia Porcellus) Number: 15 (10 test +5 negative control) Sex: No particular gender is prescribed Initial body weight: 300 ~ 500 g

Health status: healthy, not previously used in other experimental procedures

Housing: animals were housed in groups in cages identified by a card indicating the lab number,

test code and first treatment date, etc.

Animal identification: Stain with dyeing liquid

Cages: plastic cage

Acclimation period: 7 days under the same conditions as for the actual test

## 7 Animal Care and Maintenance

Animal source: Suzhou Experimental Animal Sci-Tech Co., Ltd. <Permit Code: SCXK (SU)

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2015-0007>

Bedding: corncob, Suzhou Shuangshi Laboratory Animal Feed Science Co., Ltd.

Feed: guinea pig diet, Suzhou Experimental Animal Sci-Tech Co., Ltd.

Water: drinking water met the Standards for Drinking Water Quality GB 5749-2006

Animal room temperature: 18 ~ 26°C

Animal room relative humidity: 30% ~ 70%

Lights: 12 h light/dark cycle, full-spectrum lighting

Personnel: associates involved were appropriately qualified and trained Selection: only healthy, previously unused animals were selected

There were no known contaminants present in the feed, water, or bedding expected to interfere with the test data.

## 8 Justification of Test System and Route of Administration

The albino guinea pig has been used historically for sensitization studies (Magnusson and Kligman, 1970). The guinea pig is believed to be the most sensitive animal model for this type of study. The susceptibility of the guinea pig to a known sensitizing agent, 2,4-dinitrochlorobenzene (DNCB) has been substantiated at SDWH (listed in **Table 1** and **Table 2**).

The test article was extracted and administered in vivo through a medium compatible with the test system. Dermal application corresponds to the likely route of human exposure.

## 9 Experimental Design

## 9.1 Preparation of Extracts

#### 9.1.1 Pretreatment

No pretreatment required.

#### 9.1.2 Extraction

Under aseptic conditions, samples were taken according to the sampling method (Mix the four components by equal quantity and extract together (see the image in Annex 2), Total surface area data of the test article provided by the sponsor, 568 cm<sup>2</sup>), and extracted in closed inert containers according to the extraction ratio of 3 cm<sup>2</sup>: 1 mL (sample: extraction vehicle). The extraction vehicle was 0.9% Sodium Chloride Injection (SC).

Ī	Test Period	A atual Campling	Extract Procedure			Final
	rest reriou	Actual Sampling	<b>Extract Ratio</b>	SC	Condition	Extract
V	Intradermal	Surface area	2	190.2 mI	37°C, 72 h	Not
	Induction Phase I	568 cm <sup>2</sup>	$3 \text{ cm}^2$ : $1 \text{ mL}$	189.3 IIIL	37 C, 72 II	Clear
	<b>Topical Induction</b>	Surface area	2 2 . 1 I	189.3 mL	37°C, 72 h	Not
	Phase II	$568 \text{ cm}^2$	$3 \text{ cm}^2$ : $1 \text{ mL}$	169.5 IIIL	37 C, 72 II	Clear
	Challenge Phase	Surface area	$3 \text{ cm}^2: 1 \text{ mL}$	189.3 mL	37°C, 72 h	Not
	Chancinge Phase	$568 \text{ cm}^2$	3 cm <sup>2</sup> : 1 mL	107.3 IIIL	31 C, 12 II	Clear

The extraction solvent was colorless and transparent pre- extraction and changed into slight rustiness suspension post-extraction. The extract was stored at 4°C, and tested within 24 h, without the process of adjusting its pH value, filtering, centrifuging, diluting, etc.

The vehicle (without the test article) was similarly prepared to serve as the control.

## 9.2 Experimental Procedure

#### 9.2.1 Animal Preparation and Grouping

On the first day of treatment, 15 guinea pigs were weighed and identified. The fur from the dorsoscapular area of the animals was removed with an electric clipper. Grouping as follow:

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Group Name	Group Size	Gender
Test	10 animals	No particular gender is prescribed.
Negative Control	5 animals	No particular gender is prescribed.

#### 9.2.2 Intradermal Induction Phase I

A pair of 0.1 mL intradermal injections was made for each of the following, into each animal, at the injection sites (A, B and C) as shown in Figure 1 in the clipped intrascapular region.

Site A: A 50:50 (V/V) stable emulsion of Freund's complete adjuvant mixed with the chosen solvent.

Site B: the test sample (undiluted extract); the control animals were injected with the solvent alone. Site C: the test sample at the concentration used at site B, emulsified in a  $50:50 \ (V/V)$  stable emulsion of Freund's complete adjuvant and the solvent (50%); the control animals were injected with an emulsion of the blank liquid with adjuvant.

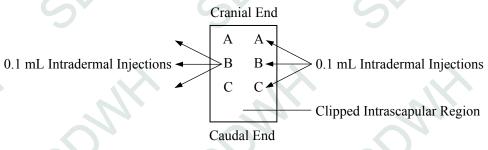


Figure 1 Locations of intradermal injection sites

### 9.2.3 Topical Induction Phase II

The maximum concentration that can be achieved in Intradermal induction phase I did not produce irritation. Animals are pretreated with 10% sodium dodecyl sulfate (Solvent: Distilled water, Date prepared: 2019-09-30) ( $24 \pm 2$ ) h before the topical induction application.

At 7 d after completion of the intradermal induction phase, administer 0.5 mL test article extract by topical application to the intrascapular region of each animal, using a patch of area approximately 8 cm<sup>2</sup> (absorbent gauze), so as to cover the intradermal injection sites. Secure the patches with an occlusive dressing. Remove the dressings and patches after  $(48 \pm 2)$  h.

Treat the control animals similarly, using the blank liquid alone.

## 9.2.4 Challenge Phase

At  $14 \pm 1$  d after completion of the topical induction phase, challenge all test and control animals with the test sample. Absorbent gauzes (2.5 cm  $\times$  2.5 cm) were soaked respectively with 0.5 mL test article and 0.5 mL control article. Apply the test article extract and control article topically to two sites that were not treated during the induction stage. Secure with an occlusive dressing. Remove the dressings and patches after (24  $\pm$  2) h.

#### 9.3 Observation of Animals

Observe the appearance of the challenge skin sites of the test and control animals  $(24 \pm 2)$  h and  $(48 \pm 2)$  h after removal of the dressings. Full-spectrum lighting was used to visualize the skin reactions. Describe and grade the skin reactions for erythema and oedema according to the Magnusson and Kligman grading given in the following table for each challenge site and at each time interval.

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Patch Test Reaction	Grading Scale
No visible change	0
Discrete or patchy erythema	1
Moderate and confluent erythema	2
Intense erythema and/or swelling	3

#### 9.4 Evaluation of Results

Magnusson and Kligman grades of 1 or greater in the test group generally indicate sensitization, provided grades of less than 1 are seen in control animals.

If grades of 1 or greater are noted in control animals, then the reactions of test animals which exceed the most severe reaction in control animals are presumed to be due to sensitization.

If the response is equivocal, rechallenge is recommended to confirm the results from the first challenge.

The outcome of the test is presented as the frequency of positive challenge results in test and control animals.

## 10 Results

The results of skin reaction after challenge were listed in **Table 3**. No skin sensitization reaction was found in the skin of guinea pigs using extracts of the test article, and the positive rate of sensitization was 0%.

The positive rate of sensitization in the positive control group was 100%, listed in **Table 1**. Clinical observations and weight changes of guinea pigs were listed in **Table 4**.

#### 11 Conclusion

Under the conditions of this study, the test article Nerve and Muscle Stimulator extract showed no significant evidence of causing skin sensitization in the guinea pig.

## 12 Record Storage

All raw data pertaining to this study and a copy of the final report are to be retained in designated SDWH archive.

## 13 Confidentiality Agreement

Statements of confidentiality were as agreed upon prior to study initiation.

## 14 Deviation Statement

There were no deviations from the approved study protocol which were judged to have any impact on the validity of the data.

## Annex 1 Test Data

Table 1 Guinea pig sensitization dermal reactions of positive control

Group	Be Animal		(24 ± 2) h Before Phase II Patch Application		(24 ± 2) h Following Challenge Phase		± 2) h lowing nge Phase	Positive Rate after Challenge
		Left	Right	Test Sites	Control Sites	Test Sites	Control Sites	Phase
	1	2	2	2	0	2	0	
Positive	2	3	2	1	0	2	0	
Control	3	2	3	2	0	3	0	100%
Control	4	2	2	3	0	3	0	
	5	3	3	2	0	2	0	
	6	0	0	0	0	0	0	
NI4:	7	0	0	0	0	0	0	
Negative	8	0	0	0	0	0	0	-
Control	9	0	0	0	0	0	0	
	10	0	0	0	0	0	0	

Note: the data of positive control come from SDWH- M201905116-1 (Completed Date: 2019-12-26)

Table 2 Weigh change and clinical observation of positive control

	Animal _	Wei	ght (g)	Clinical Observation Event
Group	Animal — Number	Before Injection	After Experiment	Clinical Observation Except Dermal Reactions
	1	326	398	Normal
Dogitivo	2	344	428	Normal
Positive	3	353	436	Normal
Control	4	343	429	Normal
	5	339	415	Normal
	6	354	438	Normal
Magativa	7	327	396	Normal
Negative	8	322	390	Normal
Control	9	352	442	Normal
	10	336	419	Normal

Note: the data of positive control come from SDWH- M201905116-1 (Completed Date: 2019-12-26)

 Table 3
 Guinea pig sensitization dermal reactions

		Table 3	Guinca	org serisi	tization ucin	iai reactic	113	
Group	Animal Number	Before	2) h Phase atch cation	Fo	4 ± 2) h llowing enge Phase	Foll	± 2) h owing ige Phase	Positive Rate after Challenge
		Left	Right	Test Sites	Control Sites	Test Sites	Control Sites	Phase
	1	0	0	0	0	0	0	
	2	0	0	0	0	0	0	
	3	0	0	0	0	0	0	
	4	0	0	0	0	0	0	
Toot	5	0	0	0	0	0	0	00/
Test	6	0	0	0	0	0	0	0%
	7	0	0	0	0	0	0	
	8	0	0	0	0	0	0	
	9	0	0	0	0	0	0	
	10	0	0	0	0	0	0	
	11	0	0	0	0	0	0	
Magativa	12	0	0	0	0	0	0	
Negative	13	0	0	0	0	0	0	-
Control	14	0	0	0	0	0	0	
	15	0	0	0	0	0	0	

 Table 4
 Weigh change and clinical observation

Group	Animal - Number	Weight (g)		Clinical Observation Event
		Before Injection	After Experiment	- Clinical Observation Except Dermal Reactions
Test	1	318	392	Normal
	2	350	433	Normal
	3	322	398	Normal
	4	323	399	Normal
	5	358	442	Normal
	6	336	410	Normal
	7	306	369	Normal
	8	353	441	Normal
	9	332	403	Normal
	10	320	393	Normal
Negative Control	11	325	401	Normal
	12	303	370	Normal
	13	351	438	Normal
	14	357	449	Normal
	15	319	394	Normal

## **Annex 2** Photograph of Test Article



## **Information Provided by Sponsor** Annex 3

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## **Production Process**

Not supplied by sponsor.

# **2 Other Information**Not supplied by sponsor.